

(12) UK Patent Application (19) GB (11) 2 298 577 (13) A

(43) Date of A Publication 11.09.1996

(21) Application No 9504758.5

(22) Date of Filing 09.03.1995

(71) Applicant(s)

University of Bristol

(Incorporated in the United Kingdom)

Senate House, Tyndall Avenue, Clifton, BRISTOL,
BS8 1TH, United Kingdom

(72) Inventor(s)

Gianni Angelini

(74) Agent and/or Address for Service

Haseltine Lake & Co
Hazlitt House, 28 Southampton Buildings,
Chancery Lane, LONDON, WC2A 1AT,
United Kingdom

(51) INT CL⁶

A61F 2/06

(52) UK CL (Edition O)

A5R RAR

(56) Documents Cited

WO 95/35072 A2 WO 95/15130 A1 WO 94/13224 A1
WO 93/21860 A1 US 4743251 A

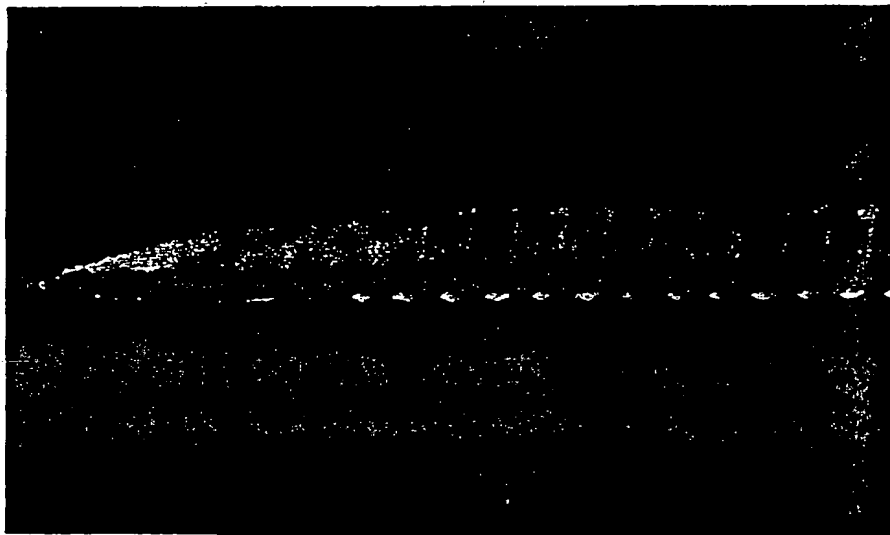
(58) Field of Search

UK CL (Edition O) A5R RAP RAR
INT CL⁶ A61B 17/11 , A61F 2/06
Online:WPI

(54) Stented vein segment for use in bypass grafting

(57) A vein segment for use in an arteriovenous bypass grafting procedure is externally stented by the provision of a non-restrictive porous stent about its outer peripheral surface. The stent may comprise a 6mm diameter polyester lock-knit tube externally supported with helically wound polypropylene, as shown in Figure 1. Alternatively, the stent may be formed from stainless steel, PTFE or biological material such as needled collagen.

FIG.1.



At least one drawing originally filed was informal and the print reproduced here is taken from a later filed formal copy.

The claims were filed later than the filing date within the period prescribed by Rule 25(1) of the Patents Rules 1995

Best Available Copy

GB 2 298 577 A

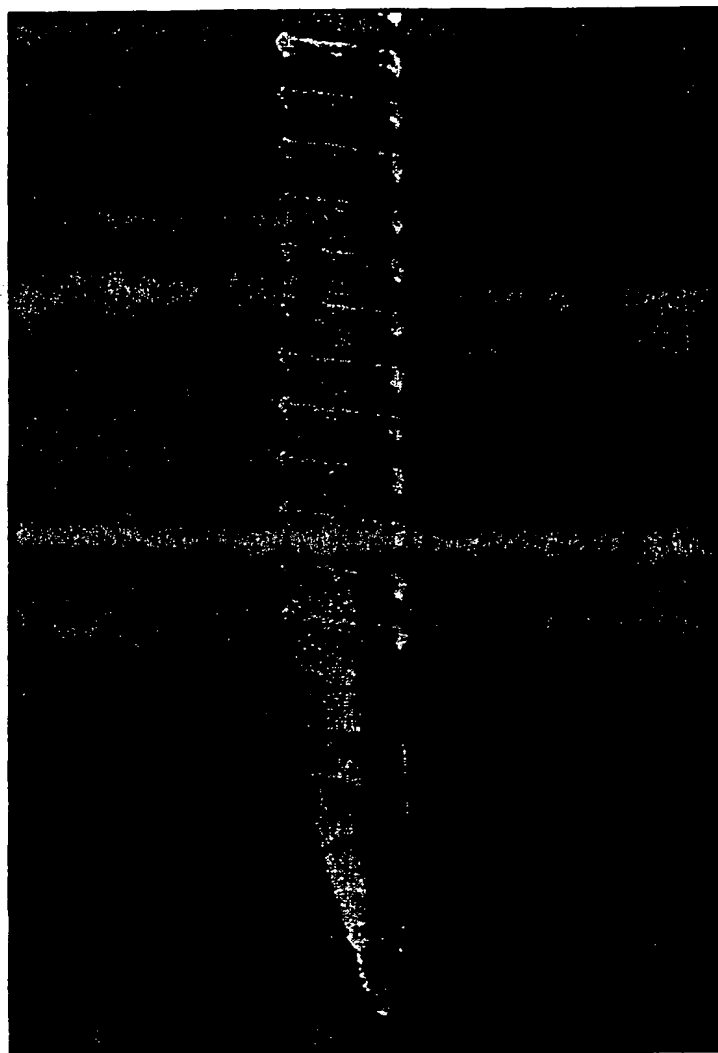


FIG.1.

FIG.2a.

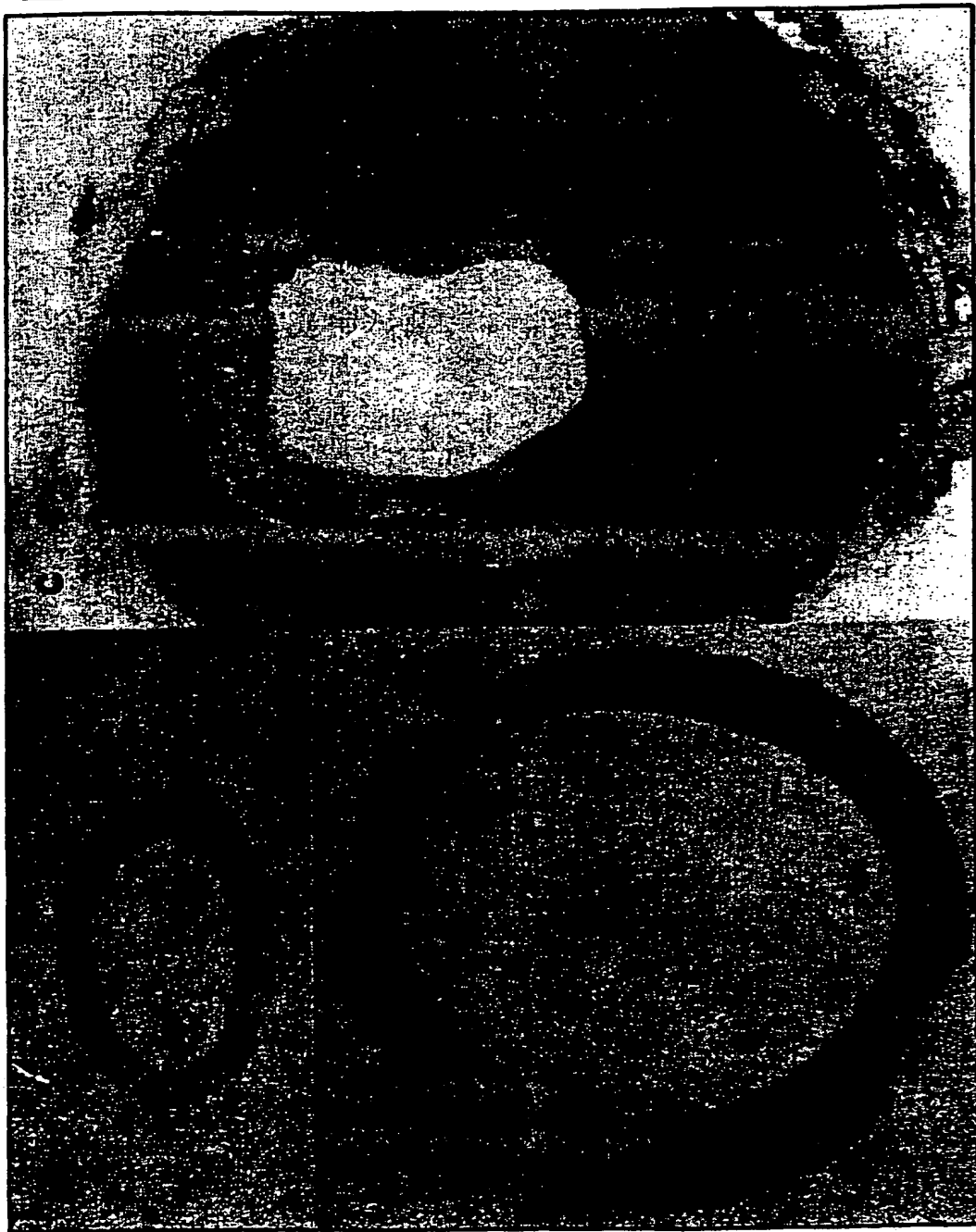


FIG.2b.

FIG.2c.

FIG.3c.

3/3

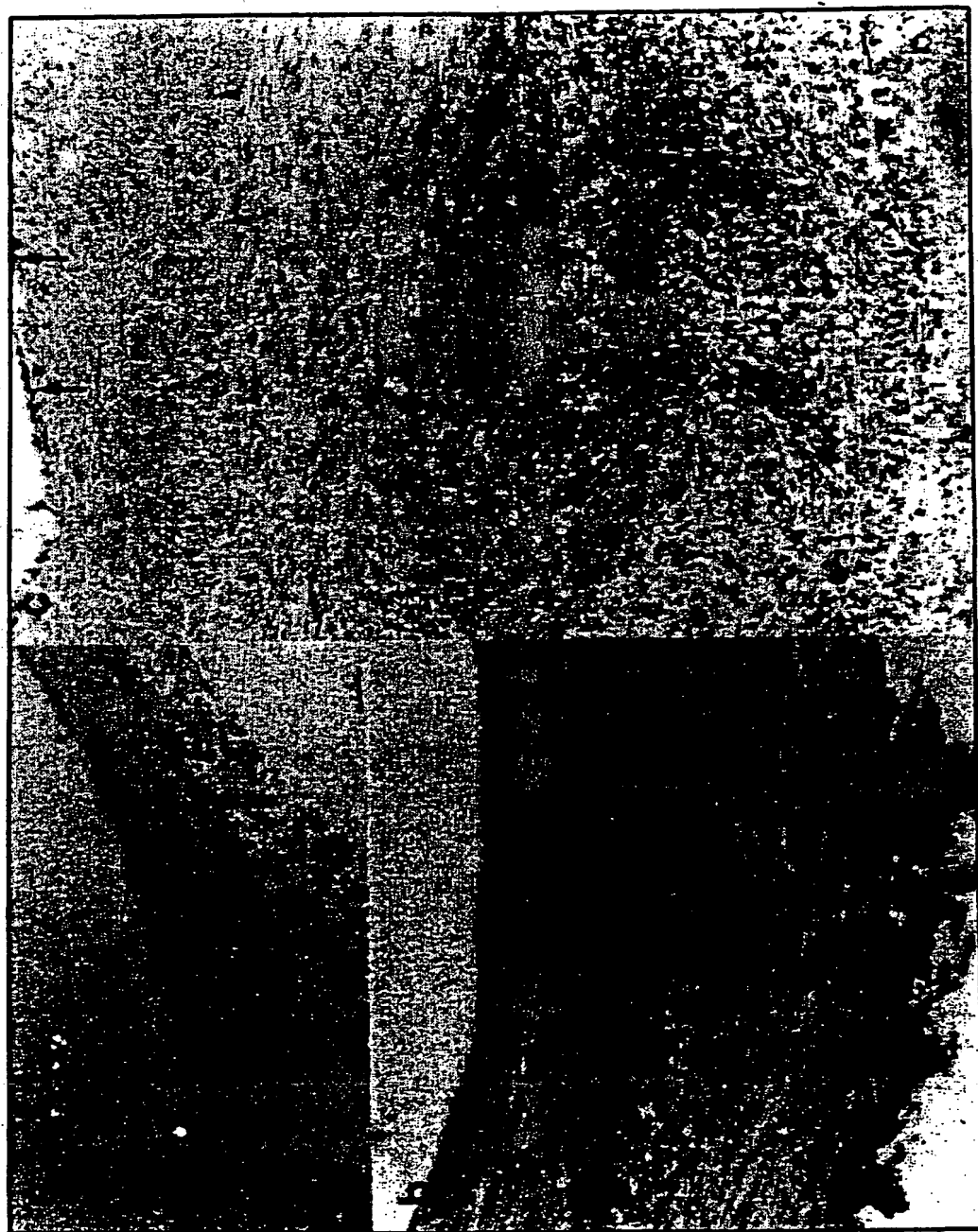


FIG.3a.

FIG.3b.

ARTERIOVENOUS BYPASS GRAFTING

This invention relates to an improvement in arteriovenous grafting.

Bypass of stenotic coronary arteries with autologous saphenous vein has become an established treatment for end-stage atherosclerotic disease, with more than 400,000 procedures in the United States alone each year. The long-term clinical success of coronary bypass grafting is limited, however, by occlusion of up to 50% of grafts within 10 years. Late occlusion results from progressive medial and neointimal thickening with superimposed vein graft atherosclerosis. Apart from lipid lowering therapy, no pharmacological or surgical intervention has been shown conclusively to influence the evolution of these changes either in man or in experimental models.

According to a first aspect of this invention, there is provided an arteriovenous bypass grafting procedure in which a vein segment is implanted into the arterial circulation of a mammalian subject, wherein a non-restrictive porous stent is provided around the grafted vein.

According to a second aspect of this invention, there is provided an externally stented vein segment for use in an arteriovenous bypass grafting procedure, said vein segment having provided about its outer peripheral surface a non-restrictive porous stent.

In the arteriovenous bypass grafting procedure of the present invention, the vein segment is implanted into the arterial circulation of a mammalian subject. This will include the coronary artery, as well as peripheral arteries such as the carotid and femoral arteries. The present invention is particularly suited to bypass of the coronary artery.

Synthetic vascular graft tubing is well known and many different constructions based on a variety of

materials are available. Typically, such graft tubing is knitted from a polyester yarn such as poly(ethylene terephthalate), sometimes referred to as Dacron. In the known surgical procedures, the synthetic vascular tubing is normally used to bypass a section of diseased vein to maintain blood flow to a patient's extremities and lower limbs.

In the present invention, vascular graft tubing is used in an entirely new way to provide an improvement in arteriovenous bypass grafting procedure. Specifically, it has been found that a porous non-restrictive external stent provided about the bypass vein segment has beneficial effects on the luminal size and the degree of medial and intimal thickening and cell proliferation in a pig vein graft model.

The porous stent used in the invention is normally of synthetic origin, but it could alternatively be formed from a biological material, such as collagen, which would have the advantage that, after a period of time, the stent would be reabsorbed into the body. If a biological material is used, it may be necessary to provide the material with the desired degree of porosity, for example by a technique such as needling.

When the porous graft tubing employed is synthetic, it will typically be of a knitted construction. The synthetic material from which the graft tubing may be formed is preferably a polyester such as poly(ethylene terephthalate), but other synthetic materials would be suitable; indeed any material which is used for implant purposes, and which can be fashioned into a porous tube, is theoretically usable, such as stainless steel, PTFE (high porosity) membrane and fibres, and other metals and polymers.

It is important that the graft tubing used as a stent in this invention is porous, in the sense that it has an ability to be invaded by cells. An indication

of porosity can be obtained by using a water permeability test which refers to the rate of flow of water through the wall of the dry prosthesis, and which provides an index of the interstitial leakage rate of the wall. A suitable such test is described in "Practical Considerations in Fabric Vascular Grafts" by B.F. Buxton et al, Am. J. Surg. 125:288-293 (1973) in which the dry prosthesis is subjected to an applied head of water of 120mm Hg, and the volume of water that passes through the wall per minute is measured. The water permeability of the stent in accordance with this test should preferably be at least 5 ml/min/cm². The preferred maximum water permeability is about 20000 ml/min/cm². For a typical synthetic grafting material such as Dacron, the water permeability would normally lie in the range of from 150 to 4000 ml/min/cm². However, for different materials, the relationship between water permeability and their porosity in terms of their use in the present invention varies, and so it is important that, for any given material, experiments are conducted to determine the ideal porosity or water permeability for use in the invention.

In the invention, it is important that the synthetic stent provided externally of the vein segment to be grafted is non-restrictive so as to allow unrestricted expansion of the graft in initial response to arterial pressure. After this initial expansion, it is also believed to be important that the internal diameter of the stent is slightly larger (for example a few mm) than the diameter of the expanded vein. The actual diameter of the stent relative to the vein must be determined empirically bearing these factors in mind, but typically the inner diameter of the stent will be at least about 3mm larger than the overall outer diameter of the vein segment to be grafted. Normally, the inner diameter of the stent will be no

more than about 6mm larger than the overall outer diameter of the expanded vein graft.

In the surgical procedure of the invention, the vein segment about which the non-restrictive stent is provided is implanted in the normal way in the artery which is to be bypassed. The stent, which is adapted in length to the vein segment to be grafted, is stitched lightly in place. The stent is left in place after the procedure, and becomes incorporated within the growing tissue which surrounds the vein.

Reference will now be made to the following drawings in which:

Figure 1 shows a graft stent for use in the present invention;

Figure 2 shows the histological appearance of (a) an ungrafted vein, (b) a representative unstented graft in a pig model, and (c) a representative stented graft in a pig model; and

Figures 3 illustrates the immunocytochemistry for proliferating cell nuclear antigen in respect of (a) an ungrafted vein, (b) a representative unstented graft in a pig model, and (c) a representative stented graft in a pig model.

White Land Race pigs (n=9, weighing 20 to 25 kg) were subjected to premedication, anaesthesia and autologous saphenous vein into common carotid artery bypass grafting by a modification of the method described previously. In brief, segments of saphenous vein were dissected using a 'no-touch' technique, rinsed in isoosmotic sodium chloride solution (0.9 g.L⁻¹) containing 2 IU/mL heparin and 50 µg/mL glyceryl trinitrate and stored in the same solution at room temperature (23°C) until needed. Both common carotid arteries of the pigs were exposed. A 3 cm segment of one artery at a time was removed and replaced with a segment of saphenous vein cut sufficiently to allow

implantation without longitudinal stretching.

Anastomoses were performed end-to-end with reversed vein, bevelled at 45° using continuous 7-0 prolene sutures. Proximal anastomoses were always performed first; in the case of stented grafts, using a stent as shown in Figure 1, the vein segments were passed through the stent before completing the distal anastomosis. The stent shown in Figure 1 comprises a continuous 6 mm diameter VP1200 polyester locknit tube externally supported with helically wound 0.8 mm polypropylene and is available from Vascutek Limited, Inchinnan, Renfrewshire, UK. The stent was cut to a slightly greater length than the graft and held in position by two single 7-0 prolene stitches placed in the adventitia of the artery at each end. The entire procedure took 50 minutes with the order of implantation of stented and unstented grafts randomised between pigs. The use of the distal or proximal portions of the veins for stented or unstented grafts was also randomised.

The animals were allowed to recover and fed a normal chow diet of 4 weeks. The grafts were then removed, pressure fixed ex-vivo at 100 mmHg for 10 mins using 10% buffered formal saline and then post-fixed in the same solution for approximately 24 hours and then processed for wax embedding.

Figure 2 illustrates the histological appearance of ungrafted saphenous vein, unstented and stented grafts. Transverse (5 µm) sections were stained with alcian blue Miller's elastin van Gieson stain and examined under light microscopy:

- a) an ungrafted vein;
- b) a representative stented graft;
- c) a representative unstented graft.

Small arrows indicate the position of the internal elastic lamina and large arrows the external elastic

lamina. The scale bar relates to all panels and represents 0.5 mm.

Figure 3 illustrates immunocytochemistry for proliferating cell nuclear antigen

- 5 a) an ungrafted vein.
- b) a representative unstented graft. Note the presence of PCNA positive cells (small arrows) in the neointima close to the luminal surface.
- 10 c) a representative stented graft. Note the presence of vasa vasorum (large arrows).

Small triangles indicate the position of the internal elastic lamina and large triangles the external elastic lamina. The scale bar relates to all
15 panels and represents 0.5 mm.

Proliferating cell nuclear antigen was detected by immunocytochemistry as described previously. Briefly, a primary monoclonal antibody (PC10, Dako Ltd, High Wycombe, Bucks, HP13 5RE, UK) was used at a 1/100
20 dilution. This was followed by a 1/50 dilution of biotinylated anti-mouse IgG (Dako) and avidin-biotin-peroxidase conjugate (Dako) according to the manufacturers instructions. Sections were counterstained with Harris' haematoxylin. Strongly
25 positive cells were counted in 5 fields per section using a x40 objective.

The following table shows the results obtained in terms of the various dimensions of saphenous vein, unstented and stented grafts:

Parameter	Ungrafted vein	Non-stented graft	Stented graft	p stented vs. non-stented
Total cross-sectional area mm ²	3.7±1.2	18.0±5.8	13.9±5.9	0.1
Luminal area mm ²	2.5±1.2	7.6±3.4	11.2±6.2	<0.05
Medial area mm ²	1.2±0.9	6.55±2.62	1.62±0.52	<0.001
Intimal area mm ²	0	3.84±3.3	1.06±0.37	<0.001
Intimal encroachment (%)	0	33.2±19.1	13.3±13.4	<0.005
Total wall thickness mm	0.13±0.06	0.85±0.38	0.25±0.14	<0.001
Medial thickness mm	0.13±0.06	0.49±0.22	0.14±0.08	<0.001
Intimal thickness mm	0	0.35±0.24	0.10±0.07	<0.001

Transverse sections obtained as described in the legend to Fig. 2 were analysed by computer-aided planimetry (MicroScale TM/TC image analysis system, Digithurst Ltd, Royston, Herts, UK), as described previously. Briefly, lumenal, intimal and medial perimeters and areas were computed using the lumenal boundary, internal and external elastic laminae as delimiters. Average intimal, medial and vessel wall thickness were derived from the area and perimeter data. Values are expressed as mean \pm S.D. and were compared using the Mann-Whitney test.

As described above segments of autologous saphenous vein were implanted into the common carotid arteries of pigs. A porous polyester stent (as shown in Figure 1) was placed around one graft while the contralateral graft served as a control. The lumenal diameter of pig saphenous vein measured *in situ* before implantation by echocardiographic ultrasonography was 1.6 ± 0.1 mm (SEM, $n=9$). Immediately after grafting into the common carotid artery the diameter of stented or unstented veins increased to 3.6 ± 0.2 mm. This was approximately 2 mm smaller than the internal diameter of the stent.

All grafts whether stented or not were patent 4 weeks after implantation. As illustrated in Figure 2 and summarised in the Table, stented and unstented grafts had a similar total (lumen plus wall) cross-sectional area that was approximately 4 times greater than that of the original vein. This confirmed that the stent allowed expansion of the graft in response to arterial pressure. Unstented grafts showed an increase in luminal area compared to ungrafted vein. However, medial enlargement, fragmentation of the internal elastic lamina and the development of a neointima also occurred (Figure 2, Table), in agreement with previous observations. Stented grafts had a significantly

greater final luminal area than unstented grafts because there was no medial enlargement compared to ungrafted vein and neointima formation was dramatically reduced almost 4-fold compared to unstented grafts. As
5 a result, the encroachment of the intima into the lumen was reduced by stenting from 33 to 13% (Fig. 2, Table).

The presence of cells progressing through the cell cycle was detected by immunocytochemistry for proliferating cell nuclear antigen (PCNA). PCNA
10 positive cells were rarely detected in ungrafted vein but were abundant in the media and the most luminal aspect of the neointima of unstented grafts (Figure 3). The medial PCNA index was reduced by stenting from $21 \pm 4\%$ to $2.4 \pm 2.2\%$ ($n=9$, $p<0.001$). The neointimal PCNA
15 index was also reduced by stenting from $24 \pm 4\%$ to $7 \pm 3\%$ ($p<0.01$). PCNA labelling in the neoadventitia was observed in both stented and unstented grafts (Fig 3). The presence of microvessels penetrating the media of
stented grafts can also be noted from Figure 3. There
20 were endothelial cells lining these vessels, as confirmed by staining for *Dolichos Bifluoros* lectin (results not shown). Such vessels were absent from the media of ungrafted vein or unstented grafts.

CLAIMS:

1. An externally stented vein segment for use in an arteriovenous bypass grafting procedure, said vein segment having provided about its outer peripheral
5 surface a non-restrictive porous stent.
2. An externally stented vein segment according to claim 1, wherein said porous stent is of synthetic origin.
3. An externally stented vein segment according
10 to claim 2, wherein the porous stent is of a knitted construction.
4. An externally stented vein segment according to claim 2 or 3, wherein the synthetic material from which the stent is formed is a polyester.
- 15 5. An externally stented vein segment according to claim 4, wherein the synthetic material from which the stent is formed is a poly(ethylene terephthalate).
6. An externally stented vein segment according to claim 1, wherein said porous stent is formed from a
20 biological material.
7. An externally stented vein segment according to any preceding claim, wherein the water permeability (as hereinbefore defined) of the stent is at least 5 ml/min/cm².
- 25 8. An externally stented vein segment according to any preceding claim, wherein the water permeability of the stent is no greater than about 20000 ml/min/cm².
9. An externally stented vein segment according to any preceding claim, wherein the water permeability
30 of the stent is in the range of from 150 to 4000 ml/min/cm².
10. An externally stented vein segment as claimed in claim 1, substantially as hereinbefore described, with reference to the accompanying drawings.



Application No: GB 9504758.5
Claims searched: 1-10

Examiner: L.V.Thomas
Date of search: 4 June 1996

Patents Act 1977
Search Report under Section 17

Databases searched:

UK Patent Office collections, including GB, EP, WO & US patent specifications, in:

UK Cl (Ed.O): A5R (RAP, RAR)

Int Cl (Ed.6): A61B 17/11, A61F 2/06

Other: ONLINE:WPI

Documents considered to be relevant:

Category	Identity of document and relevant passage	Relevant to claims
X,E	WO 95/35072 A2 (W.L.Gore) see p.3 ll.4-27	1,2
X,E	WO 95/15130 A1 (Zurbrugg) see abstract	1,2
X	WO 94/13224 A1 (W.L.Gore) see p.4 l.23 - p.5 l.14, p.5 ll.29-32, p.7 ll.1-16 & 31-34 and p.11 ll.8-10 & 14-19	1,2,4-6
A	WO 93/21860 A1 (Intervascular Inc.) see p.5 l.5 - p.6 l.11	1
A	US 4743251 (Barra) see col.1 l.62 - col.2 l.33	1

X Document indicating lack of novelty or inventive step
Y Document indicating lack of inventive step if combined with one or more other documents of same category.

& Member of the same patent family

A Document indicating technological background and/or state of the art.
P Document published on or after the declared priority date but before the filing date of this invention.
E Patent document published on or after, but with priority date earlier than, the filing date of this application.

This Page is inserted by IFW Indexing and Scanning
Operations and is not part of the Official Record

BEST AVAILABLE IMAGES

Defective images within this document are accurate representations of the original documents submitted by the applicant.

Defects in the images include but are not limited to the items checked:

- ☐ BLACK BORDERS
- ☐ IMAGE CUT OFF AT TOP, BOTTOM OR SIDES
- ☒ FADED TEXT OR DRAWING
- ☐ BLURED OR ILLEGIBLE TEXT OR DRAWING
- ☐ SKEWED/SLANTED IMAGES
- ☐ COLORED OR BLACK AND WHITE PHOTOGRAPHS
- ☐ GRAY SCALE DOCUMENTS
- ☒ LINES OR MARKS ON ORIGINAL DOCUMENT
- ☐ REPERENCE(S) OR EXHIBIT(S) SUBMITTED ARE POOR QUALITY
- ☐ OTHER: _____

IMAGES ARE BEST AVAILABLE COPY.

**As rescanning documents *will not* correct images
problems checked, please do not report the
problems to the IFW Image Problem Mailbox**